

# ***Dendrocorticiopsis orientalis* gen. et sp. nov. of the Punctulariaceae (Corticiales, Basidiomycota) revealed by molecular data**

Chia-Ling Wei<sup>1</sup>, Che-Chih Chen<sup>1,2,3</sup>, Shuang-Hui He<sup>4</sup>, Sheng-Hua Wu<sup>1,2</sup>

**1** Department of Biology, National Museum of Natural Science, Taichung 40453, Taiwan **2** Department of Plant Pathology, National Chung Hsing University, Taichung 40227, Taiwan **3** Biodiversity Research Center, Academia Sinica, Taipei 11529, Taiwan **4** School of Ecology and Nature Conservation, Beijing Forestry University, Beijing 100083, China

Corresponding authors: Shuang-Hui He ([heshuanghui@bjfu.edu.cn](mailto:heshuanghui@bjfu.edu.cn)); Sheng-Hua Wu ([shwu@mail.nmns.edu.tw](mailto:shwu@mail.nmns.edu.tw))

---

Academic editor: R. Henrik Nilsson | Received 29 March 2022 | Accepted 8 May 2022 | Published 31 May 2022

---

**Citation:** Wei C-L, Chen C-C, He S-H, Wu S-H (2022) *Dendrocorticiopsis orientalis* gen. et sp. nov. of the Punctulariaceae (Corticiales, Basidiomycota) revealed by molecular data. MycoKeys 90: 19–30. <https://doi.org/10.3897/mycokeys.90.84562>

---

## **Abstract**

*Dendrocorticiopsis orientalis* is presented in this study as a new genus and new species based on morphological and phylogenetic evidence. This new taxon is characterized by resupinate, smooth and membranaceous basidiomata, monomitic hyphal system with clamps, colorless dendrohyphidia, variable presence of cystidia, and ellipsoid to ovoid basidiospores measuring  $5\text{--}7 \times 3.2\text{--}5.2 \mu\text{m}$ . The phylogenetic analyses based on the ITS1-5.8S-ITS2 (ITS) + nuclear 28S rDNA (28S) dataset of Corticiales indicated that the new taxon is nested in Punctulariaceae, separated from other genera with strong support values. Descriptions, specimen photo, and illustrations of the new taxon are provided in this study. A morphological comparison of the four genera of Punctulariaceae is given.

## **Keywords**

Corticoid fungi, East Asia, phylogeny, taxonomy, wood-decaying fungi

## **Introduction**

Corticiales K.H. Larss. is a small order of corticioid fungi with four families: Corticiaceae Herter, Dendrominiaceae Ghobad-Nejhad, Punctulariaceae Donk, and Vuilleminiaceae Maire ex Lotsy. The members of the order show a variety of nutritional

ecologies, including lignicolous saprobes, foliicolous species, plant pathogens, and lichenicolous species (Ghobad-Nejhad et al. 2010, 2021). Species of Punctulariaceae are mainly saprobic on angiosperm trees, causing white rot. Morphologically, they are characterized by having effused to effused-reflexed basidiomata, smooth to tuberculate hymenial surface, a monomitic hyphal system with clamped generative hyphae, mostly absence of cystidia, sparsely to regularly branched dendrohyphidia, and ellipsoid to subglobose basidiospores which are negative in Melzer's reagent and acyanophilous in cotton blue. When Donk (1964) established this family, he adopted Talbot's suggestion (Talbot 1958) and designated *Punctularia* Pat. as the type genus. Ghobad-Nejhad et al. (2010) were the first to use a phylogenetic approach to analyze Punctulariaceae, and they recognized three genera, viz., *Dendrocorticium* M.J. Larsen & Gilb., *Punctularia*, and *Punctulariopsis* Ghobad-Nejhad. This arrangement was generally accepted by mycologists (Hibbett et al. 2014; He et al. 2019; Wijayawardene et al. 2020).

Most of the previous studies of Punctulariaceae focused on European species (Bernicchia and Gorjón 2010; Gorjón and Bernicchia 2017), although species from other continents received attention as well (Ghobad-Nejhad et al. 2010; Baltazar et al. 2013; Ariyawansa et al. 2015). However, the study of this family in Asia is insufficient and needs an update (Petch 1916; Cooke 1956; Guan et al. 2021). During surveys of corticioid fungi in East Asian regions, we found an unknown species morphologically similar to *Dendrocorticium* spp. Phylogenetic analyses were conducted by using ITS+28S sequences to evaluate the generic placement of the target taxon, and the results indicated that it represents a new genus and a new species of the Punctulariaceae.

## Materials and methods

### Morphological studies

Descriptions and illustrations are based on dried specimens deposited at the herbaria of the National Museum of Natural Science (TNM) and Beijing Forestry University (BJFC). Specimens were sliced into thin sections under stereo microscope (Nikon SMZ645) and mounted in 5% KOH with 1% phloxine in preparation for observations and measurements. Melzer's reagent (IKI) and cotton blue were applied to detect amyloidity or dextrinoidity, and cyanophily, respectively. Microscopic studies were carried out under 1,000× magnification using an optical microscope (Olympus BX43). For presenting the range of basidiospore dimensions, 5% values of minimum and maximum are given in parentheses.

### DNA extraction and sequencing

DNA was extracted from dried specimens using the Plant Genomic DNA Extraction Miniprep System (Viogene Biotek corporation, New Taipei City, Taiwan), following the manufacturer's protocol. ITS1-5.8S-ITS2 and partial 28S regions were amplified with the primer pairs ITS1/ITS4 (White et al. 1990) and LR0R/LR5 (Vilgalys and Hester 1990). The PCR protocols for ITS and 28S followed Chen et al. (2020). PCR

products were purified and sequenced by MB Mission Biotech company (Taipei City, Taiwan). New sequences were assembled and adjusted using BioEdit v7.2.5 (Hall 1999) and subsequently submitted to GenBank (Table 1).

**Table 1.** Information of species and strains used in phylogenetic analyses, including their localities, voucher numbers, and GenBank accession numbers (ITS and 28S). Newly generated sequences are shown in bold. Voucher number of holotypes are marked with an asterisk (\*).

Species	Locality	Voucher no.	GenBank accession no.		
			ITS	28S	
<i>Australovuilleminia coccinea</i>	Ghobad-Nejhad & Hallenb.	New Zealand	PDD:94158*	HM046875	HM046930
<i>Basidiodesertica hydei</i>		Oman	DST2020a_ SQUCC15289*	MW077150	MW077159
<i>Corticium roseum</i>		China	Ghobad-Nejhad 2428	MW805872	MW805836
<i>C. thailandicum</i>		Thailand	Ghobad-Nejhad 3012	MW805868	MW805831
<i>Cytidia salicina</i> (Fr.) Burt		Finland	Haikonen 24631	GU590881	HM046921
<i>Dendrocorticiopsis orientalis</i> Sheng H. Wu, C.L. Wei & S.H. He		Taiwan	WEI 20-166*	<b>MW580922</b>	<b>MW580924</b>
<i>D. orientalis</i>		Taiwan	WEI 20-173	<b>MW580925</b>	<b>MW580927</b>
<i>D. orientalis</i>		Taiwan	BCRC 36235	EU232219	EU232303
<i>D. orientalis</i>		China	He 4195	<b>MW580926</b>	<b>MW580921</b>
<i>Dendrocorticium polygonioides</i> (P. Karst.) M.J. Larsen & Gilb.		France	CBS 106.56	MH857525	MH869062
<i>D. roseocarneum</i> (Schwein.) M.J. Larsen & Gilb.		South Korea	KUC20121109-32	KJ668559	KJ668413
<i>Dendrominia dryina</i> (Pers.) Ghobad-Nejhad & Duhem		France	Duhem 5283	JX892936	JX892937
<i>D. ericae</i> (Duhem) Ghobad-Nejhad & Duhem		France	Duhem 4840*	JX892938	JX892939
<i>Disporotrichum dimorphosporum</i>		USA	CBS 433.85	MH861895	MH873584
<i>D. dimorphosporum</i>		Netherlands	CBS 419.70*	MH859776	MH871538
<i>Erythrimum hypnophilum</i>		France	MG169	MW805858	MW805823
<i>E. laetum</i>		—	Kotiranta 21287	GU590875	GU590878
<i>Gloeophyllum abietinum</i> (Bull.) P. Karst.		Switzerland	H 22988	JX524619	KC782733
<i>L. fuciformis</i>		Netherlands	CBS 182.49	MH856485	MH868023
<i>L. roseipellis</i>		—	CBS 299.82	EU622846	EU622844
‘ <i>Lawreymyces palicei</i> ’		—	Palice 4369*	AY542865	AY542865
‘ <i>Lawreymyces palicei</i> ’		—	Palice 2509	AY542864	AY542864
<i>Marchandiomyces aurantioroseus</i> (P. Karst.) Ghobad-Nejhad		Sweden	Hallenberg 8186	KP864659	HM046929
<i>M. corallinus</i>		—	JL128-98	AY583327	AY583331
<i>Mycobernardia incrustans</i>		France	Duhem 3613	MW805860	MW805825
<i>M. incrustans</i>		Canada	CBS172.36	MH855759	MH867272
<i>Punctularia atropurpurascens</i> (Berk. & Broome) Petch		Taiwan	WEI 17-662	<b>MW570883</b>	<b>MW570888</b>
<i>P. bambusicola</i> C.L. Zhao		China	CLZhao 9098*	MW559983	MW559985
<i>P. strigosozonata</i> (Schwein.) P.H.B. Talbot		—	HHB-11897-sp	DQ398958	AF518642
<i>Punctulariopsis efibulata</i> (M.J. Larsen & Nakasone) Ghobad-Nejhad		USA	Burdsall 8824*	KR494276	KR494277
<i>P. obducens</i> (Hjortstam & Ryvarden) Ghobad-Nejhad		Ethiopia	Ryvarden 28131	HM046918	HM046933
<i>P. subglobispora</i> (Hallenb. & Hjortstam) Ghobad-Nejhad		Argentina	Hallenberg 12761*	HM046917	HM046932
<i>Veluticeps abietina</i> (Pers.) Hjortstam & Tellería		Sweden	KHL 12474	EU118619	EU118619
<i>Vuilleminia comedens</i> (Nees) Maire		—	T-583	DQ398959	AF518666
<i>V. coryli</i> Boidin, Lanq. & Gilles		Turkmenistan	Parmasto 54999	JN387996	JN388005
<i>V. cystidiata</i> Parmasto		South Korea	KUC20131022-26	KJ668433	KJ668285
<i>V. erastii</i> Ghobad-Nejhad		Canada	DAOM 199025*	JN387998	JN388007
<i>V. macrospora</i> (Bres.) Hjortstam		France	Duhem 4860	JX892940	JX892941
<i>V. megalospora</i> Bres.		Italy	Ryvarden 43185	HM046887	HM046926
<i>V. nilsii</i> Ghobad-Nejhad & Duhem		France	Duhem 4847*	JX892947	JX892948
<i>V. pseudocystidiata</i> Boidin, Lanq. & Gilles		France	Boidin 14838*	HM046888	HM046928
<i>Waitea circinata</i>		USA	CBS472.82	MH861518	MH873265
<i>W. guianensis</i>		French Guiana	GUY13-110	MW449090	MW449101

## Phylogenetic analyses

The selection of species and samples for the ITS+28S dataset was inspired by Ghobad-Nejhad and Duhem (2014) and Guan et al. (2021). The dataset contained 43 samples from 37 species, including 35 ingroup species from 17 genera of the four families in Corticiales and 2 outgroup species from Gloeophyllales [*Gloeophyllum abietinum* (Bull.) P. Karst. and *Veluticeps abietina* (Pers.) Hjortstam & Tellería, Table 1]. Sequences were aligned in MAFFT v.7 (Kato and Standley 2013). Partitioned phylogenetic analyses were carried out for the ITS+28S dataset based on maximum likelihood (ML) and Bayesian inference (BI) methods, using MrBayes v. 3.2.6. (Ronquist et al. 2012) and RaxML Black Box (Stamatakis 2014) at the CIPRES Science Gateway (<http://www.phylo.org/>). For the BI analysis, jModeltest 2.1.10 (Darriba et al. 2012) was first executed to estimate the best-fit substitution model based on Akaike Information Criterion (AIC). The GTR+G+I was used as the substitution model for the ITS1, ITS2 and 28S regions, while K80 was used for 5.8S region. The parameter settings for ML and BI analyses followed Wu et al. (2018). Only the phylogram inferred from the ML analysis is shown since the BI and ML analyses produced similar topologies. The statistical support values are presented above the branches of the ML tree when bootstrap values (BS)  $\geq 70$  and BI posterior probability (PP)  $\geq 0.9$ . The complete phylogenetic trees and alignment were submitted to TreeBASE (submission number 29602; [www.treebase.org](http://www.treebase.org)).

## Results

### Phylogenetic inference

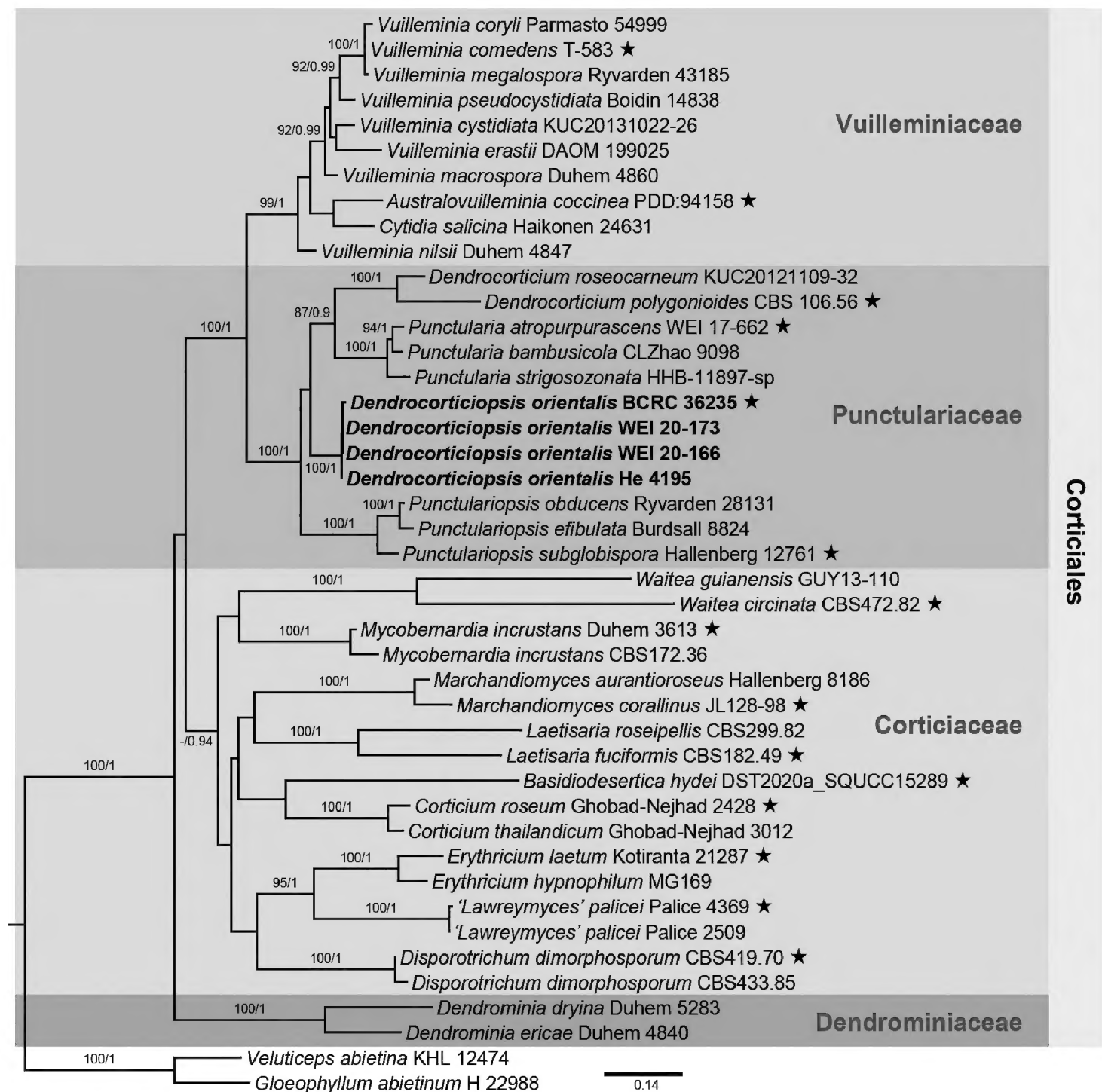
The final alignment of 43 sequences contained 1,647 sites (including gaps) of which 724 sites were from the ITS region and 923 sites from the 28S gene. Totally, 565 (34%) sites were parsimony informative. The ML tree (Fig. 1) shows the four highly supported families also recovered in previous studies (Ghobad-Nejhad and Duhem 2014; Ariyawansa et al. 2015; Ghobad-Nejhad et al. 2021; Guan et al. 2021). The four samples of the new species *Dendrocorticiopsis orientalis* formed a monophyletic group in Punctulariaceae with strong support values (BS = 100%; PP = 1), well separated from the other genera, viz., *Dendrocorticium*, *Punctularia*, and *Punctulariopsis* (Fig. 1). Therefore, *Dendrocorticiopsis* is treated as the fourth genus of Punctulariaceae.

### Taxonomy

***Dendrocorticiopsis* Sheng H. Wu, C.L. Wei & S.H. He, gen. nov.**

MycoBank: MB838902

**Diagnosis.** *Dendrocorticiopsis* differs from other genera by having strictly resupinate basidiomata, ivory hymenophore, a compact texture, a monomitic hyphal system, nodose-septate hyphae, encrusted cystidia, dendrohyphidia and ellipsoid to ovoid basidiospores.



**Figure 1.** The phylogram of Corticiales inferred from ML analysis using the combined ITS+28S dataset shows the position of *Dendrocorticopsis orientalis* (shown in bold) in Punctulariaceae. Numbers above branches indicate statistical support of BS  $\geq$  70% and PP  $\geq$  0.9. Black stars (★) indicate strains of generic species.

**Description.** Basidiomata resupinate, effused, adnate, membranaceous. Hymenial surface brownish ivory, grayish ivory to lilac ivory, smooth. Hyphal system monomitic; generative hyphae nodose-septate, colorless, slightly thick- to thick-walled. Subiculum uniform, with compact texture, usually with crystal masses; hyphae fairly horizontal. Hymenial layer thickening, with compact texture, usually with oily materials, hyphae more or less vertical. Dendrohyphidia numerous, thick-walled toward base, colorless. Cystidia clavate, apically with resinous materials. Basidia clavate to subclavate, 4-sterigmata, thick-walled toward base. Basidiospores ellipsoid to ovoid, sometimes broadly ellipsoid, smooth, thin-walled or occasionally slightly thick-walled, negative in Melzer's reagent, acyanophilous.

**Type species.** *Dendrocorticopsis orientalis*.

**Etymology.** *Dendrocorticopsis* refers to the morphological resemblance to *Dendrocorticium*.



***Dendrocorticiopsis orientalis* Sheng H. Wu, C.L. Wei & S.H. He, sp. nov.**

MycoBank: MB838903

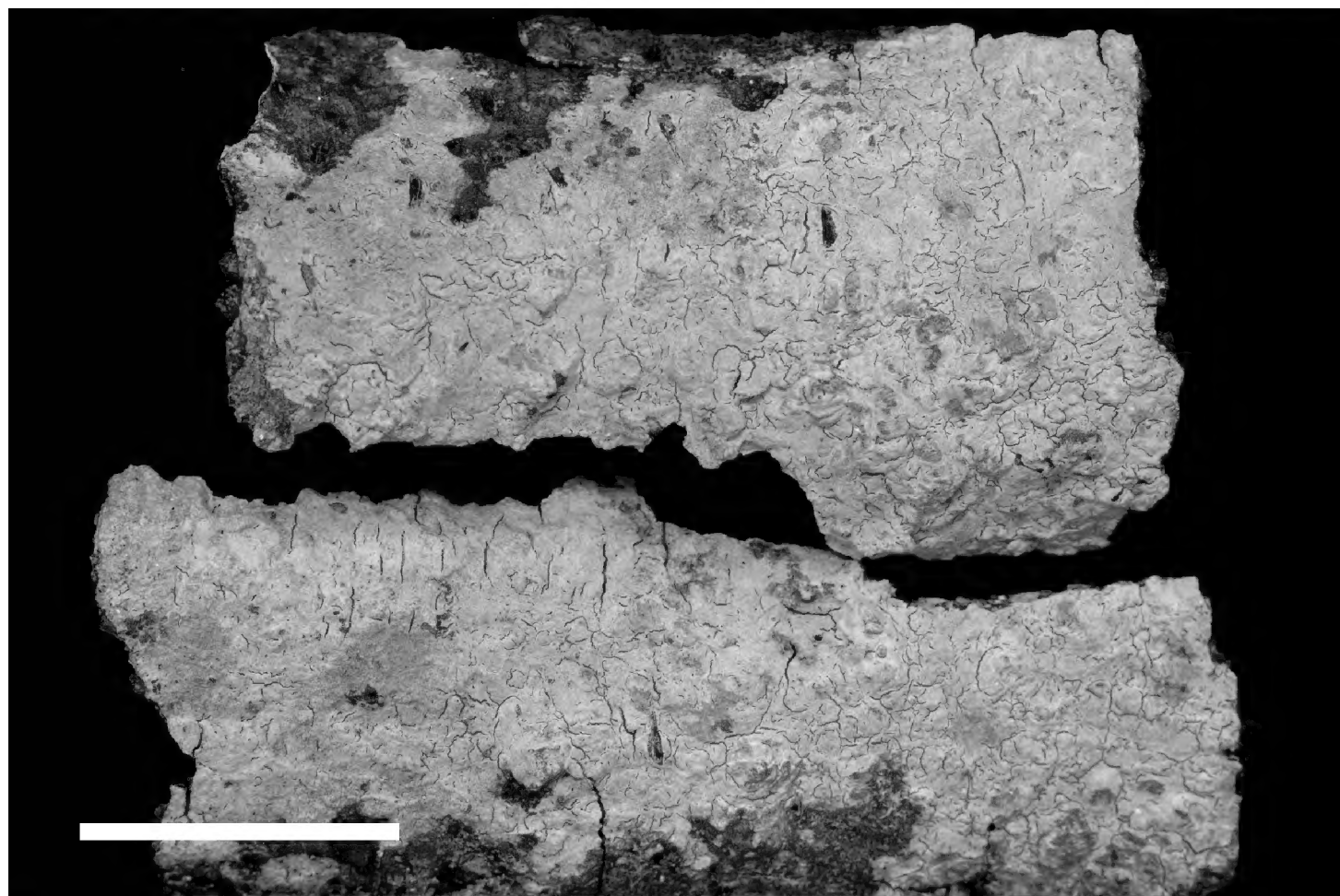
Figs 2, 3

**Diagnosis.** The noteworthy features of *Dendrocorticiopsis orientalis* are: (1) subiculum composed of a basal layer, with compact texture; (2) oily materials usually present in hymenial layer; (3) cystidia with resinous materials at apices; (4) shortly clavate to subclavate basidia; (5) ellipsoid to ovoid basidiospores measuring  $5\text{--}7 \times 3.2\text{--}5.2 \mu\text{m}$ .

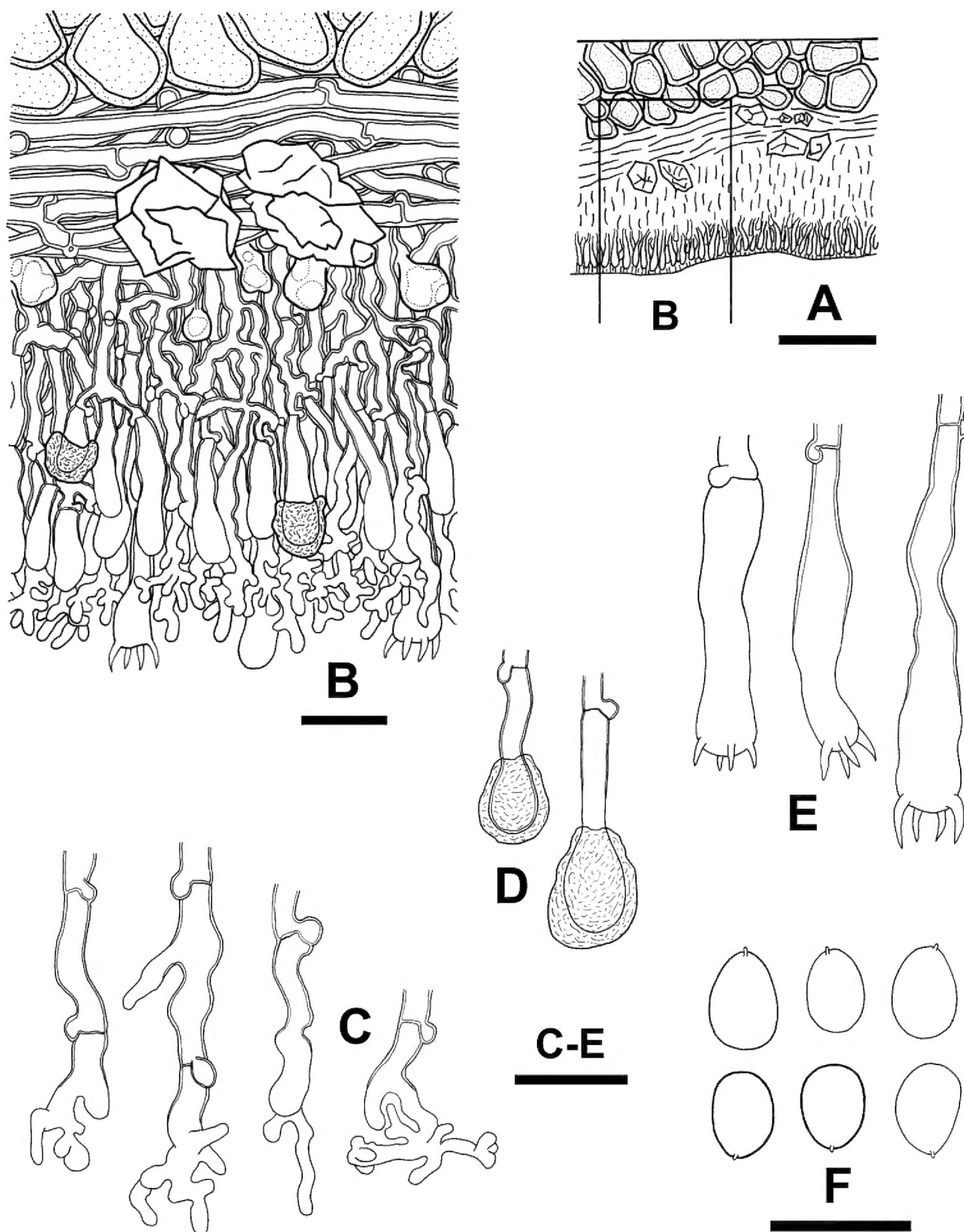
**Typification.** TAIWAN, Taichung City, Heping District, near trailhead of Mt. Tangmadan Trail,  $24^{\circ}09'53.0''\text{N}$ ,  $120^{\circ}57'26.4''\text{E}$ , 670 m asl., on dead angiosperm trunk, 20 Aug 2020, leg. C.L. Wei, WEI 20-166 (holotype, TNM F34448). GenBank: ITS = MW580922; 28S = MW580924.

**Etymology.** The epithet refers to the Eastern world, where the specimens were collected.

**Description.** Basidiomata annual, resupinate, effused, adnate, membranaceous, 50–100  $\mu\text{m}$  thick in section. Hymenial surface brownish ivory, grayish ivory to lilac ivory, smooth, finely cracked; margin concolourous, slightly pruinose, rather determinate. Hyphal system monomitic; generative hyphae nodose-septate. Subiculum fairly uniform, composed of a basal layer, with fairly compact texture, usually with crystal masses; up to 30  $\mu\text{m}$  thick, sometimes indistinct; hyphae mainly horizontal, colorless, fairly straight, 3–4  $\mu\text{m}$  diam, with walls slightly thickened up to 1  $\mu\text{m}$ . Hymenial layer thickening, with more or less compact texture, usually with oily materials, 50–70  $\mu\text{m}$



**Figure 2.** Basidiomata of *Dendrocorticiopsis orientalis* (holotype, WEI 20-166). Scale bar: 1 cm.



**Figure 3.** Micromorphological features of *Dendrocorticiopsis orientalis* (holotype, WEI 20-166) **A** profile of basidioma section **B** basidioma section **C** dendrohyphidia **D** cystidia **E** basidia **F** basidiospores. Scale bars: 50 µm (**A**); 10 µm (**B–F**).

thick; hyphae more or less vertical, colorless, 2–4 µm diam, with walls slightly thickened up to 1 µm. Dendrohyphidia numerous, 12–28 × 2–3 µm, thick-walled toward base, with walls up to 1 µm thick, colorless. Cystidia clavate, apically with resinous

materials, gradually dissolving in KOH,  $10\text{--}20 \times 3.5\text{--}5.5 \mu\text{m}$ , slightly thick-walled, or thickening toward base, with walls up to  $1 \mu\text{m}$  thick. Basidia clavate to subclavate, usually broadened at basal or middle parts,  $18\text{--}35 \times 5\text{--}7 \mu\text{m}$ , 4-sterigmata, thickening toward base, with walls up to  $1 \mu\text{m}$  thick. Basidiospores ellipsoid to ovoid, or broadly ellipsoid, smooth, colorless, with homogenous contents, thin-walled or occasionally slightly thick-walled, negative in Melzer's reagent, acyanophilous, mostly  $5\text{--}7 \times 3.2\text{--}5.2 \mu\text{m}$ .  $(5.5)6\text{--}7(7.5) \times 4.2\text{--}5.2(5.5) \mu\text{m}$ ,  $L = 6.50 \pm 0.42 \mu\text{m}$ ,  $W = 4.66 \pm 0.32 \mu\text{m}$ ,  $Q = 1.40$  ( $n = 30$ ) (holotype, WEI 20-166).  $(5.7)6.2\text{--}7(7.5) \times (4.2)4.5\text{--}5(5.2) \mu\text{m}$ ,  $L = 6.61 \pm 0.43 \mu\text{m}$ ,  $W = 4.77 \pm 0.25 \mu\text{m}$ ,  $Q = 1.39$  ( $n = 30$ ) (WEI 20-173).  $(4.2)5\text{--}6.8(7) \times (3)3.2\text{--}5(5.2) \mu\text{m}$ ,  $L = 5.8 \mu\text{m}$ ,  $W = 4.2 \mu\text{m}$ ,  $Q = 1.38$  (He 4195).

**Habitat.** On dead angiosperm wood (e.g., *Acacia* and *Castanopsis*), occurring in August.

**Distribution.** In subtropical regions, known from China: Jiangxi and Taiwan.

**Additional specimens examined (paratypes).** CHINA, Jiangxi Province, Yichun City, Yifeng County, Guanshan National Nature Reserve, 500 m asl., on dead *Castanopsis* wood, 9 Aug 2016, leg. S.H. He, He 4195 (BJFC 023637). TAIWAN, Taichung City, Heping District, near trailhead of Mt. Tangmadan Trail,  $24^{\circ}09'53.0''\text{N}$ ,  $120^{\circ}57'26.4''\text{E}$ , 670 m asl., on dead angiosperm trunk, 20 Aug 2020 leg. C.L. Wei, WEI 20-173 (TNM F0034449).

**Notes.** Both of the ITS and 28S sequences BLAST results showed that *Dendrocorticiopsis orientalis* is close to the strain BCRC 36235 that is annotated as *Ganoderma applanatum* (Pers.) Pat. in GenBank. According to personal communication with Bioresource Collection and Research Center (BCRC, Taiwan), the strain BCRC 36235 was indeed isolated from a *Ganoderma* specimen collected by Dr. Jin-Torng Peng in Nantou, Central Taiwan, on wood of *Acacia confusa* Merr. However, as suggested by Suldbold (2017), the ITS (EU232219) and 28S (EU232303) sequences of the strain BCRC 36235 are not true *G. applanatum*, and we supposed that the strain could be contaminated by *D. orientalis*, which is known to grow on *Acacia*. The specimen He 4195 collected on *Castanopsis* (Fagaceae) from Jiangxi Province has slightly smaller basidiospores ( $L = 5.8 \mu\text{m}$ ,  $W = 4.2 \mu\text{m}$ ) than the holotype.

## Discussion

A comparison of morphological characteristics for distinguishing the four genera in Punctulariaceae is provided in Table 2. *Dendrocorticiopsis* is morphologically similar to *Dendrocorticium*, however, the latter has longer and narrowly clavate to tubular basidia usually longer than  $45 \mu\text{m}$ , whereas *Dendrocorticiopsis* has clavate to subclavate basidia shorter than  $35 \mu\text{m}$ . *Punctularia* differs from *Dendrocorticiopsis* by having resupinate or effused-reflexed basidiomata with a tuberculate hymenophore, colored dendrohyphidia, and through its lack of cystidia, while *Punctulariopsis* can be distinguished from *Dendrocorticiopsis* by possessing longer basidia and basidiospores, and mostly lacking cystidia.



**Table 2.** Morphological characteristics used for distinguishing the four genera in Punctulariaceae.

	<i>Dendrocorticiopsis</i>	<i>Dendrocorticium</i>	<i>Punctularia</i>	<i>Punctulariopsis</i>
<b>basidiomata</b>	resupinate	resupinate or effused-reflexed	resupinate or effused-reflexed	resupinate
<b>hymenial surface</b>	smooth	smooth	tuberculate	smooth
<b>dendrohyphidia</b>	colourless	mostly colourless (yellowish in <i>D. roseolum</i> ); some species with encrustations	yellowish to brown or pink to rose	colourless
<b>cystidia</b>	clavate, apically with resinous materials	mostly absent ( <i>D. roseolum</i> with halocystidia; <i>D. piceinum</i> with leptocystidia)	absent	mostly absent ( <i>P. obducens</i> with leptocystidia)
<b>basidia</b>	clavate to subclavate; < 35 µm long	narrowly clavate to tubular; mostly > 45 µm long	narrowly clavate to tubular; 35–45 µm long	narrowly clavate to tubular; > 45 µm long
<b>basidiospores</b>	ellipsoid to ovoid; < 10 µm long	broadly ellipsoid to subglobose; usually < 10 µm long	ellipsoid; < 10 µm long	broadly ellipsoid to subglobose; > 10 µm long
<b>distributions</b>	subtropical regions	temperate or tropical regions	tropical to subtropical regions	tropical to subtropical regions

*Dendrocorticium violaceum* H.S. Jacks. ex M.J. Larsen & Gilb. and *D. polygonioides* (P. Karst.) M.J. Larsen & Gilb. have similar-sized basidiospores to *Dendrocorticiopsis orientalis* [4–6.5 × 3–5 µm in *D. violaceum*, 6–9 × 4–6 µm in *D. polygonioides* (Larsen and Gilbertson 1977)]. However, *D. violaceum* is distributed in Canada, has a reflexed basidiomata margin (closely adnate in *Dendrocorticiopsis orientalis*), and grows mainly on deciduous wood. *Dendrocorticium polygonioides* is mainly distributed in Europe and has a whitish to violaceous surface, large basidia (50–60 × 5–7 µm), and usually encrusted dendrohyphidia (Larsen and Gilbertson 1977).

Acknowledgements

This study was supported by a Grant-in-Aid for Scientific Research (no. 109-08.1-SB-18) from the Council of Agriculture, Executive Yuan, ROC and the National Natural Science Foundation of China (No. 31750001). We are grateful to Miss Shin-Yi Ke for DNA extraction and PCR works, and to Miss Siou-Zhen Chen for managing studied specimens.

References

Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B, Chethana KWT, Dai DQ, Dai YC, Daranagama DA, Jayawardena RS, Lücking R, Ghobad-Nejhad M, Niskanen T, Thambugala KM, Voigt K, Zhao RL, Li G-J, Doilom M, Boonmee S, Yang ZL, Cai Q, Cui Y-Y, Bahkali AH, Chen J, Cui BK, Chen JJ, Dayarathne MC, Dissanayake AJ, Ekanayaka AH, Hashimoto

- A, Hongsan S, Jones EBG, Larsson E, Li WJ, Li Q-R, Liu JK, Luo ZL, Maharachchikumbura SSN, Mapook A, McKenzie EHC, Norphanphoun C, Konta S, Pang KL, Perera RH, Phookamsak R, Phukhamsakda C, Pinruan U, Randrianjohany E, Singtripop C, Tanaka K, Tian CM, Tibpromma S, Abdel-Wahab MA, Wanasinghe DN, Wijayawardene NN, Zhang J-F, Zhang H, Abdel-Aziz FA, Wedin M, Westberg M, Ammirati JF, Bulgakov TS, Lima DX, Callaghan TM, Callac P, Chang C-H, Coca LF, Dal-Forno M, Dollhofer V, Fliegerová K, Greiner K, Griffith GW, Ho H-M, Hofstetter V, Jeewon R, Kang JC, Wen T-C, Kirk PM, Kytövuori I, Lawrey JD, Xing J, Li H, Liu ZY, Liu XZ, Liimatainen K, Lumbsch HT, Matsumura M, Moncada B, Nuankaew S, Parnmen S, de Azevedo Santiago ALCM, Sommai S, Song Y, de Souza CAF, de Souza-Motta CM, Su HY, Suetrong S, Wang Y, Wei S-F, Wen TC, Yuan HS, Zhou LW, Réblová M, Fournier J, Camporesi E, Luangsa-ard JJ, Tasanathai K, Khonsanit A, Thanakitpipattana D, Somrithipol S, Diederich P, Millanes AM, Common RS, Stadler M, Yan JY, Li XH, Lee HW, Nguyen TTT, Lee HB, Battistin E, Marsico O, Vizzini A, Vila J, Ercole E, Eberhardt U, Simonini G, Wen H-A, Chen X-H, Miettinen O, Spirin V, Hernawati (2015) Fungal diversity notes 111–252—Taxonomic and phylogenetic contributions to fungal taxa. *Fungal Diversity* 75(1): 27–274. <https://doi.org/10.1007/s13225-015-0346-5>
- Baltazar JM, Da Silveira RMB, Rajchenberg M (2013) *Asterostromella roseola* Bres. ex Rick is combined in *Dendrocorticium* (Corticaceae, Agaricomycetes). *Phytotaxa* 104(1): 49–52. <https://doi.org/10.11646/phytotaxa.104.1.7>
- Bernicchia A, Gorjón SP (2010) *Fungi Europaei* 12: Corticiaceae s.l. Edizioni Candusso, Lomazzo.
- Chen CC, Chen CY, Lim YW, Wu SH (2020) Phylogeny and taxonomy of *Ceriporia* and other related taxa and description of three new species. *Mycologia* 112(1): 64–82. <https://doi.org/10.1080/00275514.2019.1664097>
- Cooke WB (1956) The genus *Phlebia*. *Mycologia* 48(3): 386–405. <https://doi.org/10.1080/00275514.1956.12024546>
- Darriba D, Taboada GL, Doallo R, Posada D (2012) jModelTest 2: More models, new heuristics and parallel computing. *Nature Methods* 9(8): 772–772. <https://doi.org/10.1038/nmeth.2109>
- Donk MA (1964) A conspectus of the families of Aphyllophorales. *Persoonia* 3: 199–324.
- Ghobad-Nejhad M, Duhem B (2014) Novelities in the Corticiales: *Vuilleminia nilsii* sp. nov. and *Dendrominia* gen. nov. (Basidiomycota). *Mycological Progress* 13(1): 1–11. <https://doi.org/10.1007/s11557-012-0881-3>
- Ghobad-Nejhad M, Nilsson RH, Hallenberg N (2010) Phylogeny and taxonomy of the genus *Vuilleminia* (Basidiomycota) based on molecular and morphological evidence, with new insights into Corticiales. *Taxon* 59(5): 1519–1534. <https://doi.org/10.1002/tax.595016>
- Ghobad-Nejhad M, Langer E, Nakasone K, Diederich P, Nilsson RH, Rajchenberg M, Ginns J (2021) Digging Up the Roots: Taxonomic and Phylogenetic Disentanglements in Corticiaceae ss (Corticiales, Basidiomycota) and Evolution of Nutritional Modes. *Frontiers in Microbiology* 2320: e704802. <https://doi.org/10.3389/fmicb.2021.704802>
- Gorjón SP, Bernicchia A (2017) *Dendrocorticium pinsapineum* (Corticiales, Basidiomycota), second world distributional area in Italy. *Nova Hedwigia* 105(3–4): 341–346. [https://doi.org/10.1127/nova\\_hedwigia/2017/0415](https://doi.org/10.1127/nova_hedwigia/2017/0415)

- Guan QX, Zhao W, Zhao CL (2021) A new species of *Punctularia* (Punctulariaceae, Basidiomycota) from southwest China. *Phytotaxa* 489(3): 285–292. <https://doi.org/10.11646/phytotaxa.489.3.5>
- Hall TA (1999) BioEdit: A user-friendly biological sequence alignment editor and analysis program for windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- He MQ, Zhao RL, Hyde KD, Begerow D, Kemler M, Yurkov A, McKenzie EHC, Raspé O, Kakishima M, Sánchez-Ramírez S, Vellinga EC, Halling R, Papp V, Zmitrovich IV, Buyck B, Ertz D, Wijayawardene NN, Cui B-K, Schoutteten N, Liu X-Z, Li T-H, Yao Y-J, Zhu X-Y, Liu A-Q, Li G-J, Zhang M-Z, Ling Z-L, Cao B, Antonín V, Boekhout T, da Silva BDB, De Crop E, Decock C, Dima B, Dutta AK, Fell JW, Geml J, Ghobad-Nejhad M, Giachini AJ, Gibertoni TB, Gorjón SP, Haelewaters D, He S-H, Hodgkinson BP, Horak E, Hoshino T, Justo A, Lim YW, Menolli Jr N, Mešić A, Moncalvo J-M, Mueller GM, Nagy LG, Nilsson RH, Noordeloos M, Nuytinck J, Orihara T, Ratchadawan C, Rajchenberg M, Silva-Filho AGS, Sulzbacher MA, Tkalčec Z, Valenzuela R, Verbeken A, Vizzini A, Wartchow F, Wei T-Z, Weiß M, Zhao C-L, Kirk PM (2019) Notes, outline and divergence times of Basidiomycota. *Fungal Diversity* 99(1): 105–367. <https://doi.org/10.1007/s13225-019-00435-4>
- Hibbett DS, Bauer R, Binder M, Giachini AJ, Hosaka K, Justo A, Larsson E, Larsson KH, Lawrey JD, Miettinen O, Nagy LG, Nilsson RH, Weiss M, Thorn RG (2014) 14 Agaricomycetes. In: McLaughlin D, Spatafora J (Eds) *Systematics and evolution. Part A. The Mycota*, vol 7, 2<sup>nd</sup> edn. Springer-Verlag, Berlin, Heidelberg, 373–429. [https://doi.org/10.1007/978-3-642-55318-9\\_14](https://doi.org/10.1007/978-3-642-55318-9_14)
- Katoh K, Standley DM (2013) MAFFT Multiple sequence alignment software version 7: Improvements in performance and usability. *Molecular Biology and Evolution* 30(4): 772–780. <https://doi.org/10.1093/molbev/mst010>
- Larsen MJ, Gilbertson RL (1977) Studies in *Laeticorticium* (Aphylllophorales, Corticiaceae) and related genera. *Nordic Journal of Botany* 24: 99–121.
- Petch T (1916) Revisions of Ceylon fungi (Part IV). *Annals of the Royal Botanic Gardens (Peradeniya)* 6: 153–183.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics (Oxford, England)* 30(9): 1312–1313. <https://doi.org/10.1093/bioinformatics/btu033>
- Suldbold J (2017) Taxonomic re-evaluation of medicinal wood-decaying fungi: *Abundisporus*, *Fomitopsis*, and *Ganoderma* in Korea. Dissertation, Seoul National University.
- Talbot PHB (1958) Studies of some South African resupinate Hymenomycetes. Part II. *Bothalia* 7(1): 131–187. <https://doi.org/10.4102/abc.v7i1.1652>
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172(8): 4238–4246. <https://doi.org/10.1128/jb.172.8.4238-4246.1990>
- White TJ, Bruns T, Taylor LS (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds)

- PCR protocols: a guide to methods and application. Academic Press, San Diego, 322–315. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>
- Wijayawardene NN, Hyde KD, Al-Ani LKT, Tedersoo L, Haelewaters D, Rajeshkumar KC, Zhao RL, Aptroot A, Leontyev DV, Saxena RK, Tokarev YS, Dai DQ, Letcher PM, Stephenson SL, Ertz D, Lumbsch HT, Kukwa M, Issi IV, Madrid H, Phillips AJL, Selbmann L, Pfliegler WP, Horváth E, Bensch K, Kirk PM, Kolaříková K, Raja HA, Radek R, Papp V, Dima B, Ma J, Malosso E, Takamatsu S, Rambold G, Gannibal PB, Triebel D, Gautam AK, Avasthi S, Suetrong S, Timdal E, Fryar SC, Delgado G, Réblová M, Doilom M, Dolatabadi S, Pawłowska JZ, Humber RA, Kodsueb R, Sánchez-Castro I, Goto BT, Silva DKA, de Souza FA, Oehl F, da Silva GA, Silva IR, Błaszczowski J, Jobim K, Maia LC, Barbosa FR, Fiuza PO, Divakar PK, Shenoy BD, Castañeda-Ruiz RF, Somrithipol S, Lateef AA, Karunarathna SC, Tibpromma S, Mortimer PE, Wanasinghe DN, Phookamsak R, Xu J, Wang Y, Tian F, Alvarado P, Li DW, Kušan I, Matočec N, Mešić A, Tkalčec Z, Maharachchikumbura SSN, Papizadeh M, Heredia G, Wartchow F, Bakhshi M, Boehm E, Youssef N, Hustad VP, Lawrey JD, Santiago ALCMA, Bezerra JDP, Souza-Motta CM, Firmino AL, Tian Q, Houbraken J, Hongsanan S, Tanaka K, Dissanayake AJ, Monteiro JS, Grossart HP, Suija A, Weerakoon G, Etayo J, Tsurykau A, Vázquez V, Mungai P, Damm U, Li QR, Zhang H, Boonmee S, Lu YZ, Becerra AG, Kendrick B, Brearley FQ, Motiejūnaitė J, Sharma B, Khare R, Gaikwad S, Wijesundara DSA, Tang LZ, He MQ, Flakus A, Rodriguez-Flakus P, Zhurbenko MP, McKenzie EHC, Stadler M, Bhat DJ, Liu JK, Raza M, Jeewon R, Nassonova ES, Prieto M, Jayalal RGU, Erdoğan M, Yurkov A, Schnittler M, Shchepin ON, Novozhilov YK, Silva-Filho AGS, Gentekaki E, Liu P, Cavender JC, Kang Y, Mohammad S, Zhang LF, Xu RF, Li YM, Dayarathne MC, Ekanayaka AH, Wen TC, Deng CY, Pereira OL, Navathe S, Hawksworth DL, Fan XL, Dissanayake LS, Kuhnert E, Grossart HP, Thines M, (2020) Outline of Fungi and fungus-like taxa. *Mycosphere: Journal of Fungal Biology* 11(1): 1060–1456. <https://doi.org/10.5943/mycosphere/11/1/8>
- Wu SH, Chen CC, Wei CL (2018) Three new species of *Phanerochaete* (Polyporales, Basidiomycota). *MycoKeys* 41: 91–106. <https://doi.org/10.3897/mycokeys.41.29070>

## Supplementary material I

### Alignments to TreeBase

Authors: Chia-Ling Wei, Che-Chih Chen, Shuang-Hui He, Sheng-Hua Wu

Data type: Alignments (fas. file)

Explanation note: We have uploaded the alignments to TreeBase and here is the link and the file. <http://purl.org/phylo/treebase/phyloids/study/TB2:S29602?x-access-code=cdd27042a420e43e26dd8e62ea382799&format=html>.

Copyright notice: This dataset is made available under the Open Database License (<http://opendatacommons.org/licenses/odbl/1.0/>). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: <https://doi.org/10.3897/mycokeys.90.84562.suppl1>